



FERAL AND STRAY CATS MONITORING AND CONTROL, A PRELIMINARY GUIDELINE TOWARDS GOOD PRACTICE





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This document was published by NPCA (National Pest Control Agencies) which, until part way through 2018, provided a co-ordinating forum for agencies and stakeholders to address vertebrate animal pest control in New Zealand. In 2018 its role was transferred to the Ministry for Primary Industries under its Bionet brand.

PUBLICATIONS

Most of NPCA's publications on animal pest control were partially updated in April 2018 and transferred to the library section of the Ministry for Primary Industries' 'Bionet' online portal. The updates reflect the transfer and also acknowledge the change in the regulatory regime during 2017 and 2018, while not fully incorporating these changes in the interim, pending further reviews of the publications. Written by experienced practitioners, the main titles cover:

- best practice guidelines on controlling and monitoring vertebrate pests; and
- information about relevant regulations.

The transferred publications can be found at www.bionet.nz/library





FERAL AND STRAY CATS

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AMENDMENTS IN THIS EDITION

This April 2018 edition has been updated as part of an interim generic review of most NPCA publications. The purpose is twofold.

- » Firstly, to reflect the substantial change in the regulatory regime relating to Health and Safety and use of VTAs (Vertebrate Toxic Agents) in the workplace, which now both sit under the Health and Safety at Work Act 2015, and associated regulations.
- » Secondly, to change links to other NPCA publications and contact details now that NPCA's publications have been transferred to the BioNet portal, run by the Ministry for Primary Industries.

The full nature of the regulatory changes have NOT been fully captured here, and users are directed to the source legislation and website information provided by the various administering agencies.

This interim review is intended to be followed up more fully in due course.

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PART 1. INTRODUCTION

1.1 Purpose

This guideline was commissioned by the National Pest Control Agencies (NPCA) to provide a background overview of the pest cat problem in New Zealand, with practical guidance on how to monitor feral cat populations, and carry out feral cat control.

The primary audience is field staff and contractors responsible for designing, implementing and monitoring feral cat control programmes. Community groups, land holders and land managers who have a need to manage feral cat populations will also find the document a useful source of practical guidance.

1.1 Scope

This guideline should be considered preliminary. It is a 'living' document that will be periodically reviewed and updated in response to feedback and to the availability of new research findings.

1.2 Layout

The guideline is divided into three parts:

1. Knowing the feral and stray cat pest.

The basic biology and general habits of feral and stray cats are described to provide a basis for determining the nature and timing of monitoring and control. Impacts on primary production and native ecosystems are also discussed.

2. Monitoring.

The techniques to best inform management about the population status of cats in a given area are described and discussed. It is notoriously difficult to obtain a reliable estimate of cat abundance so this section is more an information piece than a specific set of guidelines.

3. Control.

Information and guidelines are provided on approaches for controlling feral and stray cats.

1.3 Acknowledgements

The NPCA would like to acknowledge Shaun Ogilvie, John McIlroy and Charles Eason for their work in creating this preliminary guideline. The Department of Conservation is acknowledged for publishing a number of current best practice information sheets on trapping systems for cats. Alastair Fairweather and Darren Peters are acknowledged for their work in the development of these systems, and for allowing the information sheets for each of the trapping systems to be reproduced at the end of this guideline. Feral and Stray Cats, Monitoring and Control, a Preliminary Guideline towards Best Practice, April 2018

PART 2. KNOWING THE FERAL AND STRAY CAT PEST

2.1 Definitions of Cat Categories Used in This Guideline

Domestic cat. A pet or house cat living in close connection with a household where its requirements are intentionally provided by humans. Domestic cats may still impact on native fauna by their predatory activities, but do not rely on hunting for food. Reproduction is usually manipulated by humans.

Stray cat. This is a cat that relies only partly on humans for provision of its ecological requirements. Stray cats may obtain food or shelter that has been provided intentionally or otherwise by humans. This category includes animals kept on farms for rodent control, dumped animals and cats living in urban fringe situations such as garbage dumps. Reproduction in these populations is not usually manipulated by humans.

Feral cat. This is a free-living cat that has minimal, or no reliance on humans, and which survives and reproduces in self-perpetuating populations.

Individual cats can potentially move between each category during their lifetimes.

2.2 History and Distribution of Cats in New Zealand

Cats (*Felis catus*) were deliberately introduced to New Zealand from 1769 onwards by European explorers. For example, Captain Cook gave two cats to Maori at Tolaga Bay on his first voyage to New Zealand. Other explorers, too, would have gifted or left cats in New Zealand. As ships visiting New Zealand carried cats to control on-board rat and mice infestations, some unintentional release of cats would undoubtedly also have occurred.

However, cats were not recorded as feral until at least 50 years later.¹ Feral cats probably became established in the North Island by the 1830s and in the South Island by the 1840s. By the 1860s they were reported as being numerous in the Canterbury area. After rabbits were introduced, cats spread further when farmers deliberately released them onto their properties to try and control the rabbits. Cats were also introduced to many offshore islands by sealers, whalers, farmers and others. While many of these populations have since died out or been eradicated, feral cats do remain on more than 10 of New Zealand's offshore islands.²

Worldwide, cats are kept as pets and to control rodents. They are found across virtually all latitudes, including many islands. Feral populations have often established from domestic pets. Within New Zealand, cats are widespread in the North, South and Stewart Islands. They range from fully feral, to reliant on human settlements for food, to fully domestic (see 2.1, Definitions). They are found living in a range of terrestrial habitats from sand dunes to tussocks, exotic or native forests and scrub, and from sea level to 3000 m.

¹ Gillies and Fitzgerald 2005

² Parkes & Murphy 2003

2.3 Physical characteristics

It is thought that the cat was originally domesticated from the African wild cats in Egypt more than 4000 years ago.

Within New Zealand, feral cats tend to fall into one of six distinctive coat patterns: striped tabby (the basic type), blotched tabby, black, grey, ginger and tortoiseshell, and all can have white patches on the body. The most dominant patterns are the two tabby and black coat colours.

Cats have sensitive hearing, being able to hear frequencies up to 65 kHz. They have welldeveloped night vision (with a green eyeshine). They can see colour in daylight but do not usually respond to colours. The activity patterns of cats ranges from diurnal to nocturnal, depending on season, habitat and the presence of a litter. Some can be active throughout the whole 24 hour period. Some sexual dimorphism is apparent, with female cats weighing around 70-80% of the weight of male cats.

2.4 Home Ranges

The home range of cats depends largely upon three factors – cat density, prey density and habitat type, although other factors can also affect it. Feral cats are usually solitary and sparsely distributed, with measured home ranges in excess of 200 ha. Domestic cats maintain a much smaller home range compared to feral cats. In times of prey scarcity, home range size increases as cats are forced to travel further for food resources. Cats will tolerate some overlap between home ranges, but maintain a core area that will be aggressively defended.

Kin groups will sometimes be established, especially in areas such as farm buildings or food waste dumps. These are often female kin groups, with a single male included or a number of males loosely associated with the group. Young males are driven out of the group as they near sexual maturity. Home range boundaries are marked using scent glands, claw sharpening on particular trees, spraying urine and leaving scats in conspicuous places.

While domestic cats usually bury their scats within their own territory, they often leave them unburied when further afield. More often than not, feral cats also leave their scats unburied, depositing them on clumps of grass or in conspicuous areas along tracks. Scats are usually dark in colour and generally consist of three to six round-to-elongate segments, containing the remains of whatever food has been recently consumed – fur, feathers and bones.

2.5 Food

Cats are well known predators. In mainland New Zealand, where there is a range of (introduced) mammal species present, cats feed predominantly on rabbits and rodents. Birds do make up a part of the diet and reptiles are important prey species at low latitudes. Other species preyed upon include invertebrates, frogs and fish; however, these are less common. Diet depends a lot on the cat's habitat. In forested or agricultural areas rabbits can be a dominant part of the diet, with possums, stoats, hedgehogs and carrion also comprising a small component.

On Stewart Island and offshore islands where prey availability is often limited, cat diet tends to rely heavily on rodents, especially ship rats or kiore rather than Norway rats. Juvenile Norway rats are taken but adults are often left as they can be very aggressive towards

predators. In these island locations, birds make up a much larger component of the diet compared to mainland populations. Seabirds and passerines (perching birds) commonly fall victim to cat predation and the species taken can vary considerably in size. In Central Otago and the McKenzie Basin, lizards and skinks are frequently preyed upon. Invertebrates are also taken, but usually only larger species such as weta, black field crickets and cicadas. Small or juvenile cats often eat more invertebrates and smaller prey, possibly because they have not developed the hunting skills of adult cats.

As well as being expert predators, cats are also opportunistic scavengers and will scavenge dead carcasses. They can contribute to the spread of some livestock diseases. Interestingly, healthy feral cats do not need access to drinking water as they obtain all they require from the prey they eat. However, in times of prey scarcity, or when suckling young, some drinking water is required.

2.6 Reproduction

Sexual maturity is reached at around eight to twelve months of age. The gestation period for cats is typically 65 days. Most kittens are born between spring and autumn and more than one litter per year is common. Feral cats have smaller litters compared to domestic cats, producing a maximum of five kittens, but often not all of these survive. Pregnant and lactating cats are often found from October to April or May, however some cases of pregnant or lactating feral cats found in winter have been recorded.

The kittens of feral cats reach 500g in around five to six weeks. Feral cats keep their kittens in the den where they were born until this weight is attained and then move to temporary dens, staying only a matter of days at each. Thereafter, feral kittens grow much more slowly compared to domestic kittens.

2.7 Population Densities

Population densities of feral cats vary greatly in New Zealand, as elsewhere in the world. Estimates for different habitats in New Zealand range from 0.17 to 5.6 cats per square km.³

2.8 Benefits and Negative Impacts on the New Zealand Environment

Feral cats potentially benefit New Zealand as predators on rabbits and rodents. It has been noted in many areas that rabbit and rodent numbers increase dramatically upon cat eradication; conversely, the numbers of these pest species are kept stable and relatively low in the presence of cats.

Apart from this potential benefit to the environment, feral cats mainly cause negative impacts.

Feral cats have been pinpointed as playing key roles in both the local and widespread decline of native birds and, in some cases, their extinction.⁴ This effect is particularly evident on islands, one example being the accelerated extinction of the Little Barrier Snipe on Little Barrier Island. They have also played a role in the decline of native lizards and invertebrates,⁵ in particular in the Central Otago and McKenzie Basin areas.

³ Gillies and Fitzgerald 2005

⁴ Morgan *et al.* 1995, Thomas *et al.* 1998

⁵ Thomas *et al.* 1998

Many offshore islands in New Zealand are used in recovery programmes for threatened native bird species; however, the presence of cats, regardless of density, is highly detrimental to this effort. Thus, a large amount of money and effort is spent eradicating cats as well as other introduced mammals from these environments. The cat eradication programme that occurred on Little Barrier Island took over 4 years to remove just 100 cats.⁶

KEY INFORMATION SOURCE

For more detailed information on cats in New Zealand, an invaluable resource is *The Handbook of New Zealand Mammals* 2nd edition (2005) edited by C. King.

⁶ Thomas et al. 1998

PART 3. MONITORING

IMPORTANT NOTE: It is difficult to obtain a reliable estimate of cat abundance. Therefore, this section is largely an information piece rather than a specific set of guidelines.

3.1 Why Monitor?

As cats are a key predator species in New Zealand, monitoring of feral cat populations is one component of successful cat management. A good monitoring regime can help managers protect native species or restore natural ecosystem equilibrium to an area.

However, monitoring of feral cat populations is not generally common practice in New Zealand and may not always be necessary if suitable alternative approaches are taken. This is reflected in the fact that the Department of Conservation (DoC) doesn't have a cat monitoring protocol. The cat control done by DoC is largely done for the protection of biodiversity, so the success of cat control is measured indirectly through the survival rate and population increase of, for example, native species being protected. As feral cats are a very difficult species to monitor, this approach taken by DoC makes considerable sense.

Monitoring is carried out to provide information for:

- a) assessing the need for a cat control programme, and
- b) assessing the success of any cat control work undertaken.

The reasons for undertaking cat control and the expected outcomes of such control need to be carefully considered when managing any pest cat population problem. In many cases, the approach taken by DoC, to monitor the survival of species being protected by cat removal, might be a better approach than monitoring the cats directly.

KEY INFORMATION SOURCES

The two references below are useful information sources for feral cat monitoring.

Mitchell, B., Balogh, S. (2007) *Monitoring Techniques for Vertebrate Pests – Cats*. Online at: <u>https://www.pestsmart.org.au/wp-content/uploads/2010/03/Monitoring-techniques-for-vertebrate-pests---cats.pdf</u>

Forsyth, D.M., Robley, A.J., Reddiex, B. (2005) *Review of methods used to estimate the abundance of feral cats.* Arthur Rylah Institute for Environmental Research, Department of Sustainability and Environment, Melbourne.

3.2 How to Approach Monitoring

The objective of monitoring is to measure abundance of the given species. There are two types of abundance measures – absolute and relative.

Absolute abundance is the number of animals estimated to be present, based on direct sighting or trapping of individuals, and is usually expressed as a density i.e. number of animals per area.⁷ Unfortunately, absolute abundance estimates can be expensive and may be unnecessary for a lot of pest management decisions.⁸ The expensive nature of absolute abundance measurement is particularly heightened for cats, as they are often solitary, can be nocturnal, and also display cryptic behaviour.⁹

Relative abundance measures are therefore more likely to be applicable for cat monitoring. Where sign such as tracks or scats are counted, it gives a relative index of the population density.¹⁰ It is assumed that this index is positively related to cat abundance, for example, more tracks will mean a higher density of cats. However, for non-solitary species a positive relationship may not always be the case and sometimes saturation can be reached (which is when 100% of monitoring devices give a positive result) at high abundances.¹¹ It is this reason which makes indices of relative abundance more suitable for feral cat monitoring, due to the solitary nature of many cats;¹² meaning saturation of indices by feral cats is less likely.

A range of monitoring methods are summarised below. In Australia, the two most commonly used techniques are track counts and spotlight counts.¹³

It should be noted that there are limitations to all the methods in regards to feral cat behaviour and activity pattern. Therefore, it would be a good idea to use at least two of the methods in combination to give the best chance of reliably estimating population abundance. Each method does have limitations, so careful planning, and knowledge of the survey area and home range of the cats to be surveyed is important. As summed up by Sargent *et al.* (1998), the "secretive habits of most carnivore species and the low density of most carnivore populations preclude accurate, precise, and inexpensive estimation of population size". Feral cats are no exception.

3.3 Techniques for Monitoring Feral Cat Populations

3.3.1 Spotlight Counts

Spotlight counts are a commonly used method for estimating population density or population size.¹⁴ We would suggest this type of monitoring take place at night, from half an hour after sunset to half an hour before sunrise,¹⁵ as follows:

⁷ Schwarz & Seber 1999, Forsyth et al. 2005

⁸ Warburton *et al.* 2004

⁹ Gillies & Fitzgerald 2005

¹⁰ Schwarz & Seber 1999

¹¹ Forsyth *et al.* 2005

¹² Gillies & Fitzgerald 2005

¹³ Forsyth *et al.* 2005

¹⁴ Edwards *et al.* 2000

¹⁵ Mitchell & Balogh 2007

- 1. Drive a vehicle slowly along a track or a road, with one or two observers using spotlights from either the roof or the tray of the vehicle to see cats.¹⁶
- Sweep spotlight arcs of 180° ahead of the vehicle¹⁷ to detect eye shine cat eye shine is generally green.¹⁸
- 3. Upon sighting an animal, stop the vehicle and use binoculars to confirm that it is a cat¹⁹ and estimate the distance from the vehicle.
- 4. Record sightings. A sample count sheet that can be used with this method is available in Appendix 1.
- 5. Calculate cat density as the number of individuals spotted per km² of area covered with the spotlight.

It is useful to be aware that there are a number of considerations that can influence spotlight counts of feral cats. Activity patterns of feral cats can range from diurnal to nocturnal, depending on weather conditions, season and presence of a litter;²⁰ thus, spotlighting can miss cats that are not active during the night hours.²¹ Animals may not look at the spotlight, and feral cats are not known to follow roads, which is typically the route taken by vehicles when spotlighting.²² Ability to spot feral cats is affected by terrain and vegetation cover through space and time. Feral cats are relatively small, solitary and sparsely distributed within home ranges²³ and may move away from spotlights.

3.3.2 Track Counts

This monitoring method can be used for any animal that leaves tracks or imprints; however, it is particularly effective for elusive animals such as cats.²⁴

- 1. Establish transects, generally along roads or pathways that can be accessed in any conditions. The transects should ideally cover a large proportion of the area being surveyed.
- 2. Make sand plots at least every hundred metres along the transects and mark the GPS coordinates of each on a map.
- 3. To undertake a track count, prepare a plot surface of fine sand or similar material to ensure that any tracks left by cats will have a good chance of being identified. The simplest method is to use sand, either taken to the plot site or similar material already at the site, raked to create a smooth surface. To enhance the likelihood of cat visitation, meat bait can be left in the centre of each plot.
- 4. Count and record identified cat tracks against each sand plot.
- 5. Counts should generally be repeated for three consecutive mornings, with the sand being smoothed again after each count.
- 6. Calculate cat density as the number of individuals tracked per km² of area surveyed with the tracking plots.

¹⁶ Short *et al.* 1997

¹⁷ Jones & Coman 1982, Ralls & Eberhardt 1997

¹⁸ Jones & Coman 1982

¹⁹ Ralls & Eberhardt 1997

²⁰ Gillies & Fitzgerald 2005

²¹ Edwards *et al.* 2000

²² Edwards *et al.* 2000

²³ Gillies & Fitzgerald 2005

²⁴ Mitchell & Balogh 2007

As individual cats can theoretically visit multiple plots, there could be a lot of error around densities estimated using this method. However, the method will be particularly useful for showing large reductions in cat density associated with a control programme, by comparing track counts before and after cat control (as per Part 4 below).

Monitoring using tracking is relatively easy to set up and use, comparatively inexpensive compared with other monitoring methods, ideal for solitary animals such as feral cats, and animal behaviour is not altered by detection.²⁵ Example cat footprints are shown in Figure1 below. The toe pads are arranged almost in a semicircle in front of the large central pad and claw marks are not usually visible.²⁶



Figure 1. Example cat footprints.

3.3.3 Tracking Tunnels

A variation of the above track count method is the use of tracking tunnels²⁷ for feral cats. A tunnel typically comprises a wooden or solid base, a tray and a cover. The tray is divided into three compartments – a central compartment that contains an ink pad and paper compartments on either side.²⁸ The size of the opening at either end of the tunnel needs to be large enough so cats enter, although the size of openings for many tracking tunnels is determined by what non-target species need to be excluded.²⁹

The same basic idea follows as for the track counts, whereby:

- 1. Transects are established as described above.
- 2. Tunnels are set up at a maximum of 100-metre intervals along the transects. Tunnels are usually baited with meat bait.
- 3. The species of interest enters the tunnel, crosses over the ink pad and leaves identifiable footprints on the paper.
- 4. When tunnels are checked, the tracked paper is removed and dated, and new paper is laid in its place.

²⁵ Mitchell & Balogh 2007

²⁶ Triggs 2004

²⁷ Domigan & Hughey 2008, Moylan & Hudson 2008

²⁸ King *et al.* 1994

²⁹ Domigan & Hughey 2008

5. As with the track count method, counts should be repeated at least three consecutive mornings and the cat density calculated in the same way.

Positioning of the tunnels is an important factor. The open ends of the tunnels should be protected from rain so the paper is not affected and the ink not diluted, as this can affect identification of tracks.

Benefits of this method include the relative ease to set up and use; the low cost, depending on the tunnel type used; and the ability to remove the papers and identify the tracks at a later more convenient time. The method also produces generally good results in relation to the target species.³⁰

Drawbacks to using tracking tunnels include the repeated entry of a single individual - a problem that can be exacerbated by baiting³¹- and that the large bulky tunnels are labour-intensive to set up and can require tracks to be cut to set them along.³²

3.3.4 Live Capture Trapping

Methods for live capture trapping are described in detail in Part 4.3.

- 1. Transects are established as described above.
- 2. Capture traps are set up, as described in detail in Part 4.3, at a maximum of 100metre intervals along the transects.
- 3. As with the track count method, counts should be repeated at least three consecutive mornings and the cat density calculated in the same way.

3.3.5 Mark-Recapture

The basis of this monitoring method is the capture, unique marking and release, then recapture of individuals.³³ The size of the population (N) can then be estimated from the proportion of marked animals that are recaptured, based on the following formula:

N = CM / R

Where

M = the number of individuals marked in the first capture

C = total number of individuals captured in second capture

R = number of individuals in second capture that are marked

There are various other mark-recapture models available. A relevant discussion can be found in Buckland *et al.* (2000). Some key assumptions must be met:

- a) All individuals have equal probability of capture and recapture,
- b) Capture does not affect survival rate, and
- c) Marks or tags are not lost or overlooked.

³⁰ King et al. 1994

³¹ King *et al.* 1994

³² Domigan & Hughey 2008

³³ Buckland *et al.* 2000

A drawback to mark-recapture is the time and labour intensive nature of this method. Also depending on factors such as season, the catch per unit effort can be variable.³⁴ It has also been noted that cats can be reluctant to enter enclosed spaces.³⁵ Cats may also be difficult to recapture, which would violate assumption number 1 above.

3.3.6 Scat Counts

Scat counts is another potentially useful monitoring method for feral cats.³⁶ Scats are collected and identified from size, shape and odour,³⁷ and much useful information can be obtained about diet.³⁸ It is non-invasive and does not alter animal behaviour. However, with this method, it can be difficult to find scats and then to confirm that they are cat scats. This method may therefore be most useful if conducted in parallel with another monitoring method, such as identification of species from hair, camera traps or DNA identification from scats.³⁹

3.3.7 DNA Analysis

Difficulties arising from reliable scat identification as a population abundance estimator can be overcome using molecular scatology⁴⁰ – the extraction and replication of DNA material from cells deposited with the faeces.⁴¹ DNA can also be extracted from hair samples⁴² caught on barbed wire⁴³ or glue patch traps⁴⁴ deliberately set for this purpose.

There are a number of advantages to using this method:

- Species, individual and even gender can be unequivocally identified, improving the accuracy of abundance estimates,⁴⁵
- It is a non-invasive method,⁴⁶ compared with blood or tissue samples,⁴⁷
- Animals do not need to be observed depositing the scat or hair,⁴⁸
- Comparatively small amounts of DNA are required,⁴⁹
- Setting hair capture traps is quick and easy,⁵⁰

³⁴ Mitchell & Balogh 2007

³⁵ Forsyth *et al.* 2005

³⁶ Davison et al. 2002

³⁷ Farrell et al. 2000, Davison et al. 2002, Triggs 2004

³⁸ Hansen & Jacobsen 1999

³⁹ Davison *et al.* 2002

⁴⁰ Hansen & Jacobsen 1999, Farrell *et al.* 2000

⁴¹ Davison *et al.* 2002

⁴² Mills *et al.* 2000

⁴³ Mowat & Strobeck 2000

⁴⁴ Mowat & Paetkau 2002

⁴⁵ Piggott & Taylor 2003

⁴⁶ Mitchell & Balogh 2007

⁴⁷ Hansen & Jacobsen 1999

⁴⁸ Piggott & Taylor 2003

⁴⁹ Mowat & Paetkau 2002

⁵⁰ Mowat & Strobeck 2000

Information about diet and even home range can be obtained.⁵¹

However, scat collection for feral cats is still problematic, as mentioned above. To achieve the best molecular scatology results, fresh scat (less than one week old) are required⁵² as after this, degradation occurs,⁵³ yet age of samples cannot always be controlled. Seasonal differences brought about largely by weather, humidity and rainfall can affect the quality of DNA able to be extracted from faecal collections⁵⁴ and DNA material from scats and hair is often of low quantity or quality.⁵⁵

Probably the most important consideration is that DNA analysis is highly technical and, therefore, can be expensive, especially for low quality DNA from scats. Despite all of this, molecular analyses from even degraded DNA can yield useful population monitoring information.

3.3.8 Camera Trapping

A monitoring method that has become more popular in the last decade is camera trapping,⁵⁶ where a photograph is taken when an animal triggers a sensor, usually an active or passive infra-red sensor.⁵⁷ Camera trapping has not been widely used for population abundance estimates of feral cats,⁵⁸ but rather for large game and large wild cats such as snow leopards or tigers.⁵⁹ It is now described as a mainstream tool in conservation and ecology, with uses ranging from the discovery of new species to abundance estimation.

Karanth & Nichols (1998) used a capture-recapture model based on being able to identify individuals. By contrast, Carbone *et al.* (2001) suggest that the number of days for a camera to capture a photograph is useful for estimating population density of species which are not individually identifiable. As feral cats tend to be solitary, often nocturnal and sometimes cryptic in their behaviour,⁶⁰ and individuals are not always identifiable, the use of indirect abundance estimates, such as camera trapping, can be useful - particularly in conjunction with another indirect monitoring method such as track counts, as behaviour and activity patterns are unlikely to be disturbed by these indirect methods.

Care needs to be taken in the design and set-up of camera trap monitoring, and a pilot study can be invaluable. A key assumption made when carrying out camera trapping is that all animals have the same probability of being captured, especially when using a capture-recapture technique. Thus, grid sizes must be set to encompass the smallest home range size, to avoid gaps in the data collected.⁶¹ Social structure of cat populations in the survey area needs to be considered, as feral cats can aggressively defend their territory or, alternatively, have loosely associated kin groups.⁶² Knowledge of the terrain, vegetation cover, the camera field of view and flash strength, weather conditions and length of battery

- ⁵⁶ Rowcliffe & Carbone, 2008
- 57 Jackson *et al.* 2005

⁵⁹ Karanth & Nichols 1998

⁶¹ Jackson *et al.* 2005

⁵¹ Piggott & Taylor 2003

⁵² Piggott 2004

⁵³ Davison et al. 2000

⁵⁴ Piggott 2004

⁵⁵ Piggott & Taylor 2003

⁵⁸ Forsyth *et al.* 2005

⁶⁰ Gillies & Fitzgerald 2005

⁶² Gillies & Fitzgerald 2005

life, are amongst the factors that need to be known prior to setting up a monitoring study. Nevertheless, if these factors are worked out, it appears that camera trapping could be a highly useful tool for monitoring population abundance of feral cats.

There are an increasingly large number of commercial camera traps now available, often sold as wildlife monitoring devices for monitoring game animal species such as deer. A search of the internet will quickly elucidate potential sources and pricings.

3.3.9 Bait-Take

This method involves relating the amount of bait (usually non-toxic) taken in a set study area to abundance of feral cats. This gives an index of relative abundance.⁶³ This method can also be used when a toxic baiting operation has occurred to give an estimate of kill success.⁶⁴ For example when toxic bait take reduces to near zero, it is assumed that most of the cats have been removed.

There are some problems when using bait-take as a relative abundance index. Some animals are known to cache bait, giving an overestimation of population abundance.⁶⁵ In the case of feral cats, bait may be taken to feed young cats. Burrows *et al.* (2003) used cyanide transects to estimate relative abundance of feral cats in the Gibson Desert, Western Australia. They found that bait-take by feral cats was largely influenced by season and prey availability, making it an unreliable technique.

Problems can also be encountered when species other than cats take bait,⁶⁶ again reducing reliability of this method. Animals can learn behaviours related to baits, potentially avoiding taking bait or, conversely, seeing it as an "easy" food source. Animals may take more bait per individual than is anticipated, artificially inflating estimates of population abundance and, again, decreasing reliability. Bait-take as a method for estimating feral cat population abundance does not appear to be particularly robust or reliable.

⁶³ Forsyth et al. 2005

⁶⁴ Burrows *et al.* 2003

⁶⁵ Van Polanen Petel *et al.* 2001

⁶⁶ Forsyth et al. 2005

PART 4. CONTROL

4.1 How to Approach Feral Cat Control

The toolbox of techniques for controlling stray cat populations is quite limited.

To ensure a successful outcome, a number of factors are important when planning feral cat control.

- Knowledge of the cat population and whether it is predominantly a scavenging or hunting population can influence the success of certain control methods, as can density of cats at a site, and trapping effort.⁶⁷
- Seasonal fluctuations in weather and prey abundance and, therefore, cat hunger are also important;⁶⁸ if prey is abundant, food lures may not be effective, decreasing the success of control.

Previous research⁶⁹ has shown that trapability of feral cats varies depending on whether the population being targeted is predominantly a scavenging or hunting population. Short *et al.* (2002) found that scavenging cats, i.e. those around rubbish tips, towns, workshops or associated with human dwellings, were easier to catch using cage traps compared to cats living and hunting in the bush. Leg-hold traps may be more effective for control of hard-to-catch cats than cage traps.⁷⁰

Season is an important consideration when trapping feral cats. Molsher (2001) found higher trapping rates occurred in late autumn to early winter, whereas Short *et al.* (2002) found captures were highest in the first half of the year. Weather is obviously related to season and periods of poor weather can result in poor cat catches.

4.2 Techniques for Control of Feral Cats

A number of techniques are documented as having been used for feral cat management. These include:

- cage traps,⁷¹
- leg-hold traps,72
- exclusion fencing,⁷³
- shooting,
- dogging with shooting, and
- the use of toxins, specifically 1080, within New Zealand. PAPP is more recently also available for feral cat control.

⁶⁷ Short *et al.* 2002

⁶⁸ Keedwell & Brown 2001

⁶⁹ Short et al. 2002

⁷⁰ Sharp & Saunders 2004b

⁷¹ Veitch 2001, Sharp & Saunders 2004a

⁷² Veitch 2001, Sharp & Saunders 2004b

⁷³ Sharp & Saunders 2004b

Here, we give guidelines for the use of capture traps and kill traps as these are currently the most commonly used methods for controlling feral cat populations. These methods can be enhanced with lures, such as food, olfactory or social scents, and visual lures.⁷⁴

4.3 Capture Traps

Capture traps usually capture animals live and can be divided into two broad categories, leghold traps and cage traps. Both types of capture traps are in common use for management of feral and stray cats in New Zealand.

The Animal Welfare Act 1999 requires that traps be checked daily within 12 hours of sunrise as follows⁷⁵.

36 Obligation to inspect traps

- (1) A person who, for the purpose of capturing alive a mammal, bird, reptile, or amphibian, sets a trap or causes a trap to be set must inspect that trap, or cause a competent person to inspect that trap, within 12 hours after sunrise on each day the trap remains set, beginning on the day immediately after the day on which the trap is set.
- (2) A person who, for the purpose of capturing alive a mammal, bird, reptile, or amphibian, sets a trap or causes a trap to be set must
 - (a) remove, or cause to be removed, any live animal found in that trap; or

(b) attend properly to the care of any such animal or, without delay, kill the animal.

Leg-hold traps have further restrictions described in the Animal Welfare (leg-hold traps) Order 2007 as follows.

8 Restriction on use in certain areas of all leg-hold traps from 1 January 2008

Unless the use of the trap is pursuant to and in accordance with the conditions of an approval given under <u>clause 10</u>, no person may use a leg-hold trap —

- (a) within 150 metres of any dwellinghouse (but excluding a hut on public conservation land) without the express permission of the occupier; or
- (b) in any area where there is a probable risk of catching a companion animal.

4.3.1 Leg-hold Traps

Restricted traps should not be used (Animal Welfare (leg-hold traps) Order 2007). Either No. 1 unpadded jaw or 1.5 soft catch traps coil spring traps are suitable. Supplier information for these traps can be found in Appendix 2, DOC Feral Cat Restraining System 1.

A length of elastic 'bungy' cord may be incorporated into the anchor chain () to act as a shock absorber to reduce the likelihood of the animal dislocating or fracturing its leg when caught.

⁷⁴ Molsher 2001, Short *et al.* 2002

⁷⁵ This is the wording passed in the May 2015 amendment.

The three basic variations in the way that coil spring traps can be set are described and illustrated in Appendix 2. They are:

- blind set
- double walk-through set
- set in a specially-designed tunnel such as the Scott Theobold (ST) Chimney Tunnel.

A key factor when deciding which set type to use is the likely capture of valuable non-target species. For example, the ST Chimney Tunnel would be a prudent choice in areas where kiwi are known to be present.

4.3.2 Cage Traps

Cage traps are the preferred technique for use in urban/residential areas because:

- fewer injuries are likely to result compared to leg-hold traps
- if domestic cats are trapped, they can be released unharmed
- if required, trapped feral cats can be transported away from the area to be destroyed humanely
- the Animal Welfare (leg-hold traps) Order 2007 requirements make leg-hold trapping in urban areas impractical.

Cage traps are generally made of wire mesh and have a trigger device that closes the entrance of the trap when activated by the entry of a cat. Cats are notably cautious about entering enclosed spaces, therefore a trap size suitable for cats should be selected. While there are a number of trap types commercially available, here we describe using the Havahart Model 1089 cage trap.

Supplier information and detailed instructions and illustrations for using this capture trap can be found in Appendix 3, DOC Restraining System 2 at the back of this guideline.

Identifying Captured Cats as Feral

It is important that captured cats are correctly identified as feral, especially if conducting a control operation in an urban or residential area where domestic cats are present.

- *Presence of collar*. Firstly check for a collar and any attached tags that could indicate the animal is a domestic pet.
- *Behaviour*. Cats can become very distressed and aggressive when caged, even domestic pets, although this behaviour may be more extreme in a feral cat, and domestics can be comparatively easy to handle.
- Physical form. In the absence of collars, and when aggressive distressed behaviour is being displayed, there are some differences in physical form between feral and domestic cats. Feral cats in good physical condition have overall increased muscle development, being especially noticeable around the head, neck and shoulder region. They are predominantly short-haired, with coat colour ranging across all of those seen for domestic cats ginger, tortoise shell, black, grey and tabby and, while some white markings may be present, for example on the paws or chest, completely white feral cats are very rare.

If conducting a control operation in an urban or residential area, it is advisable to notify the public and ask owners to keep domestic cats indoors during times when the cages will be open, i.e. overnight, to reduce the chance of capturing a domestic cat.

4.4 Kill Traps

Kill traps are trapping systems where a cat is lured to a meat bait and then triggers the trap, which closes over its neck, killing it by occlusion of the carotid arteries. Kill traps have the advantages of not needing to be checked daily and despatching the animal quickly. They should only be used in situations where it is unlikely that domestic cats would be trapped.

There are a number of commercially available kill traps. The following three types of trapping systems are described in the appendices:

- Belisle Super X 220 trap in an ST-type chimney tunnel.
- Elevated Steve Allen SS cat trap with an access ramp.
- Elevated Timms trap with an access ramp.

Each of these kill trapping systems has passed draft National Animal Welfare Advisory Committee (NAWAC) standards for humane kill of feral cats. Each system has also been specifically designed to reduce the likelihood of non-target kills.

Supplier information and detailed instructions and illustrations for using each of these kill trap systems can be found in Appendices 4, 5 and 6 at the back of this guideline.

4.5 A Note on Toxins

Two toxins exist for feral cat control in New Zealand.

- 0.1% 1080 Feral Cat Bait: (Animal Control Products)

 A fish meal/polymer pellet for use by the Department of Conservation only. Controlled Substances License (CSL) required.
- 2. PredaSTOP for feral cats (PAPP). CSL required. Despite limited industry experience with this product, the performance standard sheet (DOC) provides some guidance on good practice.

Toxin use subject to label requirements and other applicable legislation. For more information refer the WorkSafe <u>www.worksafe.govt.nz</u>

4.5.1 Health and Safety Considerations

Health and safety of operators should be an important consideration. Handling of caged cats can be dangerous and appropriate protective equipment should be worn i.e. heavy leather gloves, and appropriate equipment used i.e. a catching pole. All operators should have up-to-date tetanus immunisation in case of bites or scratches. Cats can carry other diseases which are transmittable to humans, such as ringworm or toxoplasmosis. Hands should be thoroughly washed after any handling of cats, carcasses or associated equipment.

For more information refer the WorkSafe www.worksafe.govt.nz

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APPENDIX 1. EXAMPLE COUNT SHEET FOR SPOTLIGHTING

Deter		0.4		Denne of				
Date:		Site:		Page: of				
Start time:		Start Odometer:		Observer:				
Finish time:		Finish Odometer: Driver:						
Spotlight power (Volt	is, Watts):	Position: Roof-mounted Sitting hand-held						
Temperature °C:		Wind: nil light medium strong Direction:						
Cloud cover %:		Moon visibility: 0 1/4 1/2 3/4 Full						
Last rain: >week ago this week yesterday today now								
Transect section	Cat (Y/N)	Distance from vehicle (m)		Comments				

APPENDIX 2. DOC FERAL CAT RESTRAINING TRAP SYSTEM 1

The *DOC Feral Cat Restraining Trap System 1* best practice information sheet is reproduced on the following pages courtesy of the copyright holder, Department of Conservation

PREDATOR TRAPS

Current Best Practice Feral Cat Restraining Trap System 1

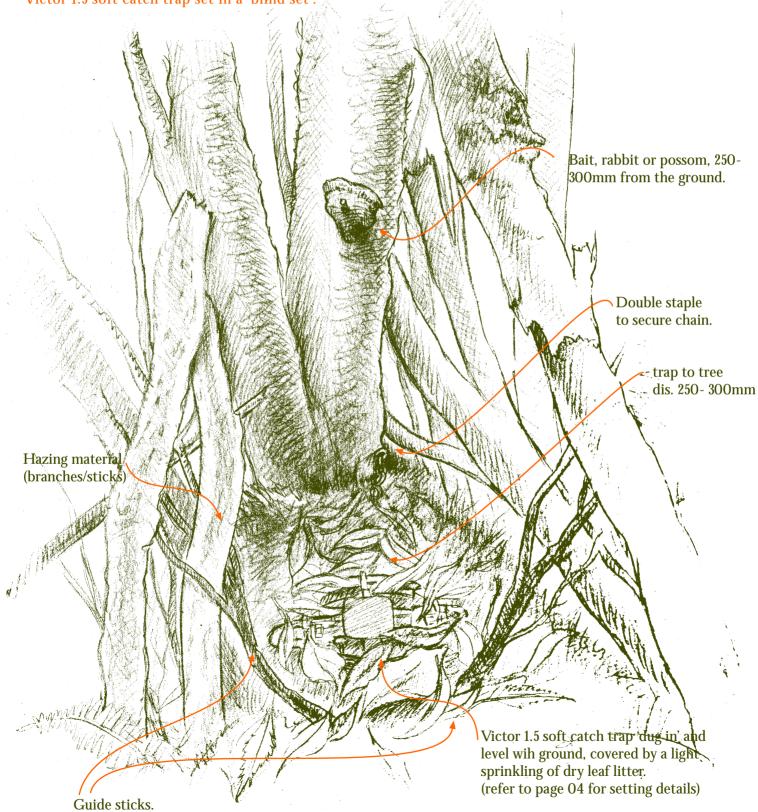


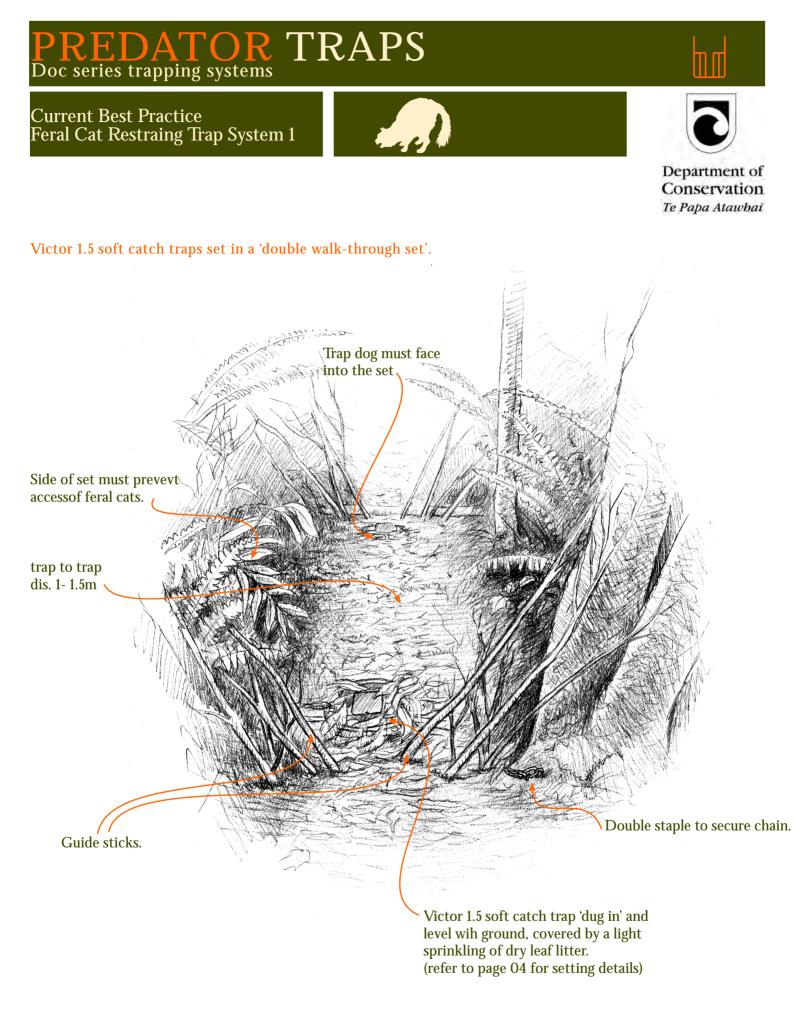
These Department of Conservation 'current best practice' set designs, must be used with Victor 1.5 soft catch traps. These sets are designed to exclude non target species, guide target species and provide public safety.

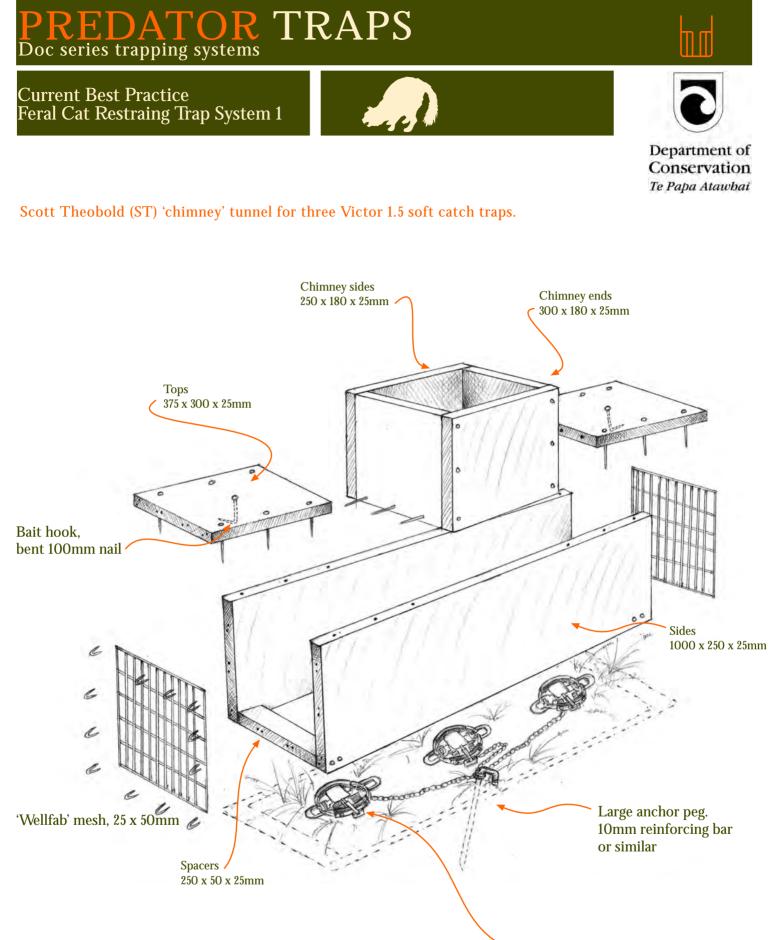
Victor 1.5 soft catch trap set in a 'blind set'.



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Three Victor 1.5 soft catch traps 'dug in' and level wih ground. (refer to page 04 for setting details)

PREDATOR TRAPS

Current Best Practice Feral Cat Restraining Trap System 1

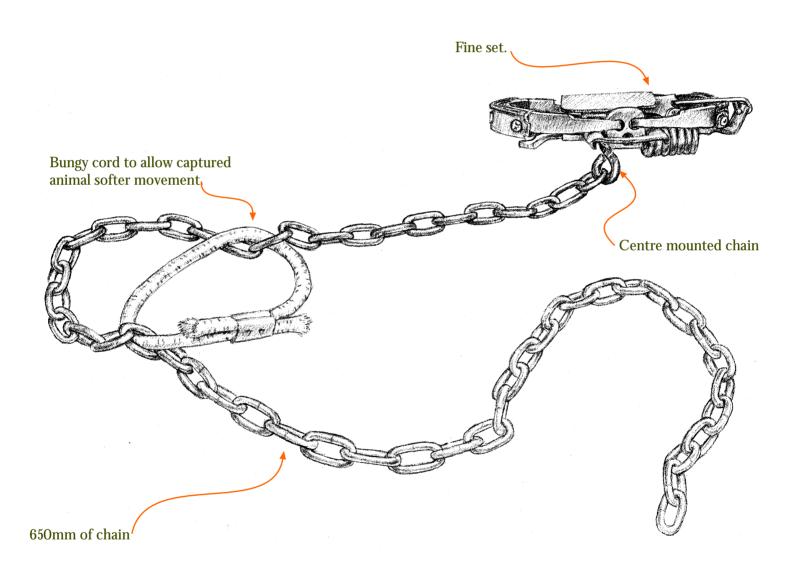




Department of Conservation Te Papa Atawbai

Victor 1.5 soft catch trap: Setting details.

All Victor 1.5 soft catch traps must be configured in the way shown here to meet DOC current best practice.



PREDATOR TRAPS

Trap Purchase/ Information

Trap Purchase

Victor 1.5 soft catch traps are available in New Zealand from:

| MS Woodcraft Ltd. 128 Marine Pde Mt Maunganui, Tauranga T| 07 575 5920 F| 07 574 8910

| Pest Management Services T| 0800 111 nopest F| 04 293 1456 E| general@nopest.co.nz A| P.O. Box 121 Waikanae. Kapiti 6454, N.Z

Wood Tunnel Purchase

Wood tunnels can be purchased from:

| Haines Pallet Co. Ltd. T| 04 568 6898 F| 04 5686480 E| haines.pallets@paradise.net.nz A| 111 Hutt Park Road, Seaview.

Advice and contacts

Predator control advice, trap development contacts and feedback.

Alastair Fairweather
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Research Development & Improvement
T | 07 8580013 F| 07 858 0001
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A | 1st Floor
Vero House
127 Alexandra Street,
PO Box 112
Hamilton

Department of Conservation National Predator Control Research Development & Improvement T| 04 471 3256 F| 04 471 3279 E| dpeters@doc.govt.nz A| P.O. Box 10-420 65 Victoria Street Wellington.



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APPENDIX 3. DOC FERAL CAT RESTRAINING TRAP SYSTEM 2

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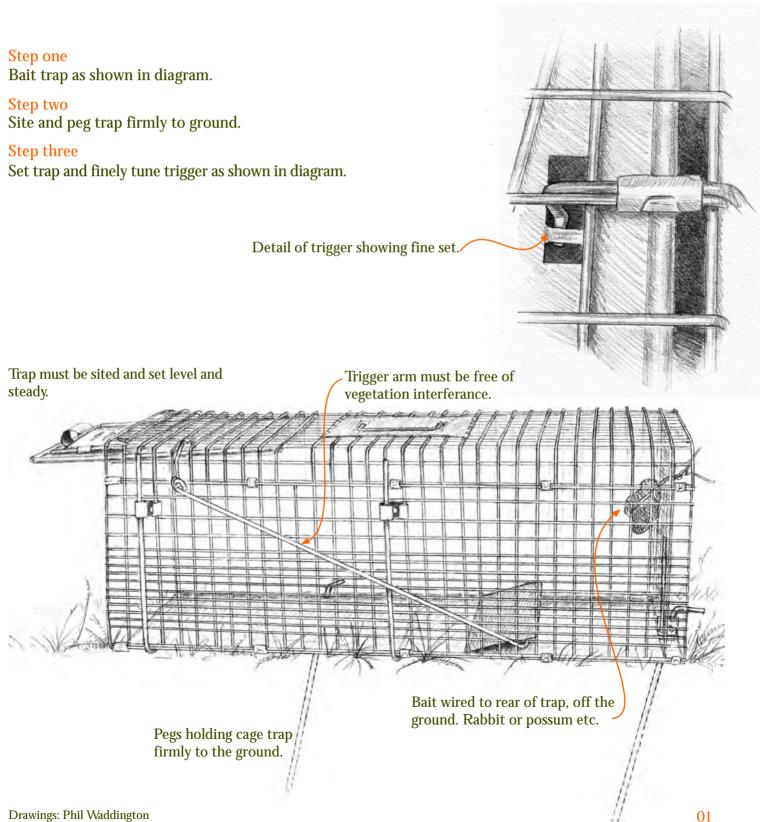
TRAPS Doc series trapping systems

Current Best Practice Feral Cat Restraining Trap System 2



This Department of Conservation 'current best practice' set design, must be used with Havahart model 1089 cage traps. This set is designed to exclude non target species, guide target species and provide public safety. These setting instructions must be followed to meet these standards.





Trap Purchase/ Information





Trap Purchase

Havahart model 1089 cage traps are available in New Zealand from:

| MS Woodcraft Ltd. 128 Marine Pde Mt Maunganui, Tauranga T| 07 575 5920 F| 07 574 8910



Department of Conservation Te Papa Atawhai

Advice and contacts

Predator control advice, trap development contacts and feedback.

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A| P.O. Box 10-420
65 Victoria Street
Wellington.

APPENDIX 4. DOC FERAL CAT KILL TRAP SYSTEM 1

The DOC Feral Cat Kill Trapping System 1 best practice information sheet is reproduced on the following pages courtesy of the copyright holder, Department of Conservation

PREDATOR TRAPS Doc series trapping systems

Current Best Practice Feral Cat Kill Trapping System 1



This Department of Conservation 'current best practice' tunnel design, must be used with Belisle Super X 220's.

This tunnel is designed to exclude non target species, guide target species and provide public safety.

The Belisle Super X 220 set in a 'chimney' tunnel has passed* draft NAWAC (National Animal Welfare Advisory Committee) standards as a humane kill trap for feral cats. These setting instructions must be followed to meet these standards.

Step one

Bait trap trigger as shown in diagram

Step two

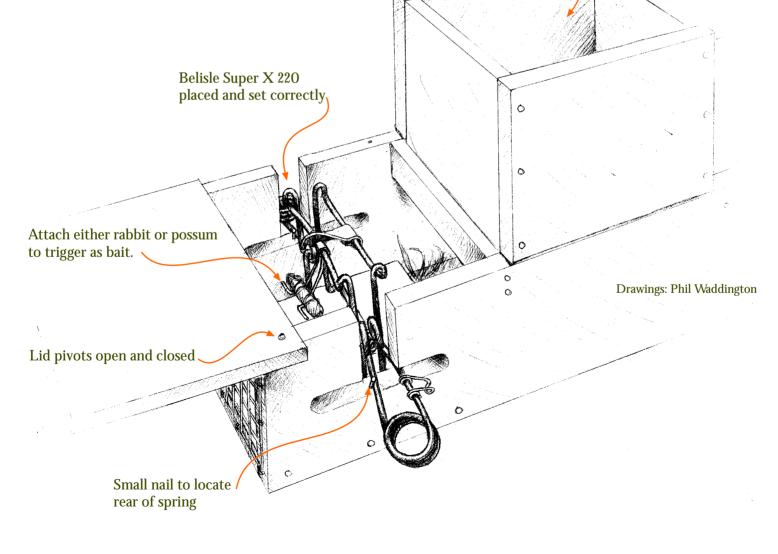
Set trap outside box (traps come with setting instructions)

Step three

Place into box as shown

Removeable chimney (lifts off for access to traps, see tunnel design sheet).





Trap Purchase/ Information





Trap Purchase

Belisle Super X 220's are available direct from Canada. They supply direct via. the internet, Head Office can assist with the purchase. | Trans Canada Trapline Co 1867 Bond street. Box 21020 North Bay Ontario P1B9N8 Canada www.trapsandfur.com

Trap Tunnel Purchase

Trap tunnels can be purchased from:

| Haines Pallet Co. Ltd. T| 04 568 6898 F| 04 5686480 E| haines.pallets@paradise.net.nz A| 111 Hutt Park Road, Seaview.

Setting Tool Purchase

Robert Rose Ltd. T| 021 711 653 E| robbie_@xtra.co.nz

Advice and contacts

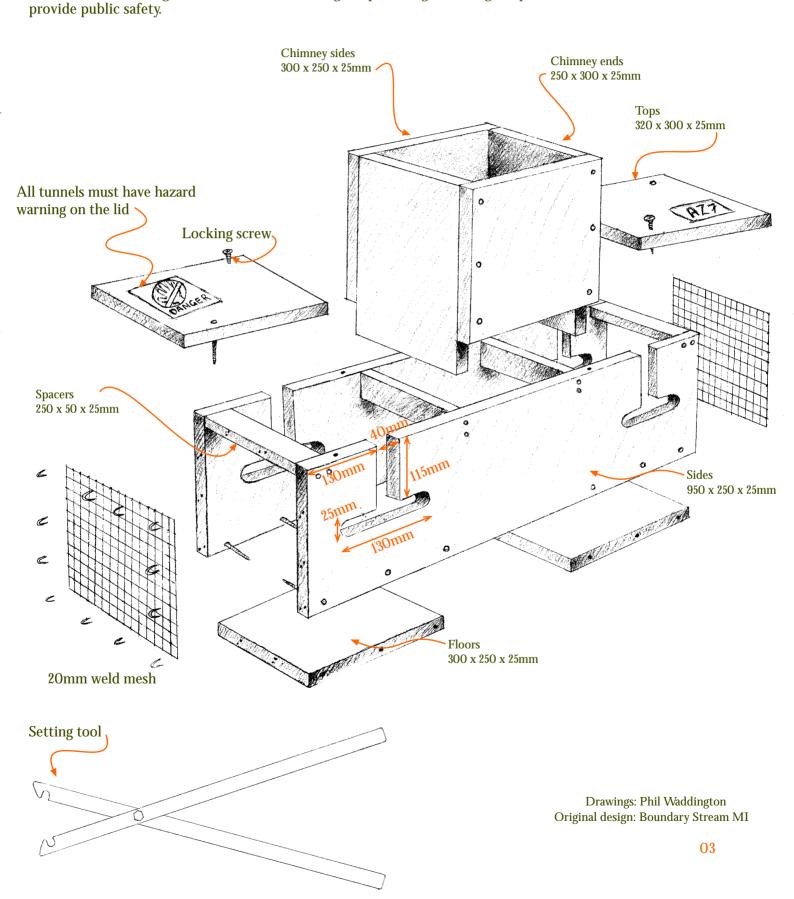
Predator control advice, trap development contacts and feedback.

Alastair FairweatherDepartment of ConservationAnimal and Plant PestsResearch Development & ImprovementT| 07 8580013F| 07 858 0001E| afairweather@doc.govt.nzA| 1st FloorVero House127 Alexandra Street,PO Box 112Hamilton



Tunnel Design

This Department of Conservation current best practice 'chimney' tunnel design, must be used with Belisle Super X 220 traps. This tunnel is designed to exclude non target species, guide target species and





APPENDIX 5. DOC FERAL CAT KILL TRAP SYSTEM 2

The *DOC Feral Cat Kill Trapping System 2* best practice information sheet is reproduced on the following pages courtesy of the copyright holder, Department of Conservation

Current Best Practice Feral Cat Kill Trapping System 2



This Department of Conservation 'current best practice' ramp/cubby design, must be used with Steve Allen (SA) conibear traps.

This ramp/cubby is designed to exclude non target species, guide target species and provide public safety.

The SA conibear trap set on a 'ramp' has passed* draft NAWAC (National Animal Welfare Advisory Committee) standards as a humane kill trap for feral cats.

These setting instructions must be followed to meet these standards.

Step one

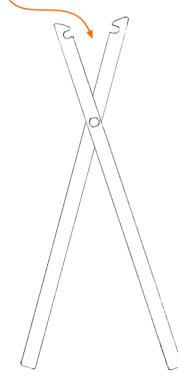
Construct, set and attach permalat mount.

Step two

Place bait onto mount, set trap and slide into mount with trap trigger uppermost and outwards as shown in diagram.

One half handful of bait (minced rabbit) is placed below and behind trap triggers.

Setting tool



Drawings: Phil Waddington

SA conibear set on 'permalat' mount, provided with trap.

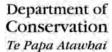
Extra bait as lure

Ramp built with 75mm x 25mm or similar timber.

Stapled chain attached to a post or tree, level with the base of the set trap. 1000mm - 1200mm above ground.

* The killing effectiveness of a modified Steve Allen Conibear trapping system for capturing feral cats (wgncr-63175)





PREDATOR TRAPS Doc series trapping systems

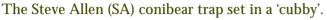
Current Best Practice Feral Cat Kill Trapping System 2



c

200mm

ē



Step one

Construct and attach cubby with permalat mount to tree or post.

Step two

Place bait onto mount, set trap and slide into mount with trap trigger uppermost and outwards as shown in diagram.

Cubby screwed or nailed to post or tree 200mm

212mm

9mm H4 treated ply or similar

One half handful of bait (minced / rabbit) is placed below and behind trap triggers.

Drawings: Phil Waddington Original design: Jimbo McConochie

Ramp built with 75mm x 25mm or similar timber.

Stapled chain attached to a post or tree, level with the base of the set trap. 1000mm - 1200mm above ground.

250mm



OR TRAPS Doc series trapping systems



Trap Purchase/ Information



Department of Conservation Te Papa Atawhai

Trap Purchase

SA Conibear traps are available direct from:

- **|** Terry Johnson
- T 09 4374573 09 4339772
- E tnjohnson@doc.govt.nz,tnjohnson@slingshot.co.nz A Mclennan Road,
 - Whangarei

Trap cubby Purchase

Trap cubbys can be purchased from: Haines Pallet Co. Ltd. T| 04 568 6898 F| 04 5686480 E haines.pallets@paradise.net.nz A 111 Hutt Park Road, Seaview.

Setting Tool Purchase

Robert Rose Ltd. T| 021 711 653 E robbie_@xtra.co.nz

Advice and contacts

Predator control advice, trap development contacts and feedback. Alastair Fairweather **Department of Conservation** Animal and Plant Pests **Research Development & Improvement** F| 07 858 0001 T| 07 8580013 afairweather@doc.govt.nz Εİ A 1st Floor Vero House 127 Alexandra Street, PO Box 112 Hamilton

Darren Peters **Department of Conservation** National Predator Control **Research Development & Improvement** F| 04 471 3279 T| 04 471 3256 E dpeters@doc.govt.nz A Level 3 **65 Victoria Street** P.O. Box 10-420 Wellington.

APPENDIX 6. DOC FERAL CAT KILL TRAP SYSTEM 3

The *DOC Feral Cat Kill Trapping System 1* best practice information sheet is reproduced on the following pages courtesy of the copyright holder, Department of Conservation

R TRAPS Doc series trapping systems

Current Best Practice Feral Cat Kill Trapping System 3



This Department of Conservation 'current best practice' set design, must be used with Timms traps.

This set is designed to exclude non target species, guide target species and provide public safety.

The Timms trap has passed* draft NAWAC (National Animal Welfare Advisory Committee) standards as a humane kill trap for feral cats. These setting instructions must be followed to meet these standards.

Step one

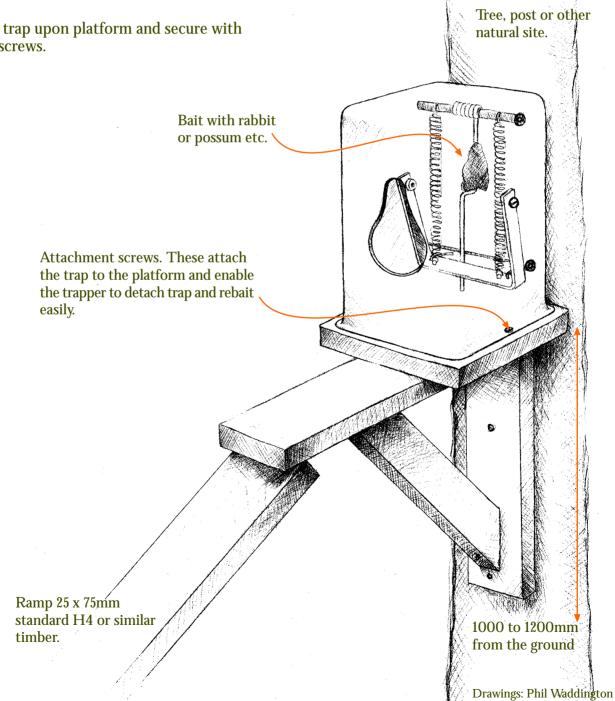
Thread bait onto trap trigger up to and past bent step, as shown in diagram.

Step two

Place baited trap upon platform and secure with attachment screws.

Step three

Set trap.



EDATOR TRAPS Doc series trapping systems

Trap Purchase/ Information

Trap Purchase

Timms traps are available in New Zealand from: **K.B.L** Rotational Moulders P.O. Box 827 Palmerston North T| 06 358 6477 F| 06 355 4825 E sales@kbl.co.nz



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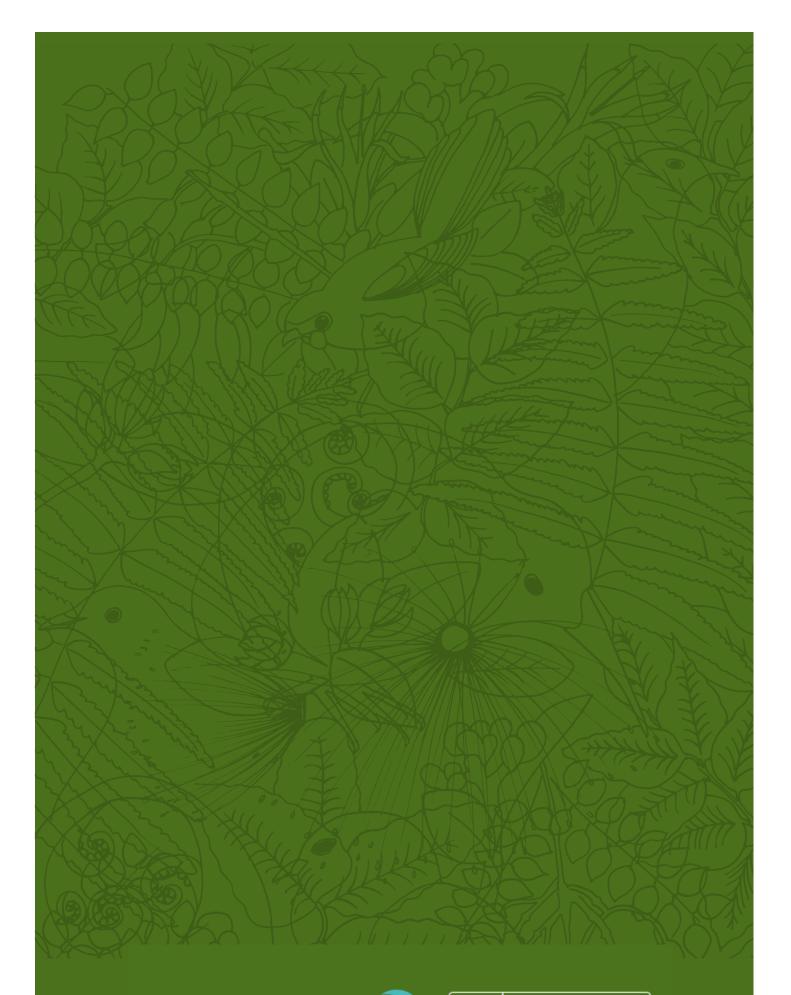
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